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Synthesis, spectral characterization and biological evaluation of 4*H*-1,4-benzothiazines, their sulfones and ribofuranosides

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1. Introduction

The synthesis of benzothiazines, their sulfones and ribofuranosides has attracted tremendous interest due to wide spectrum of biological activities possessed by these compounds such as antibacterial, CNS depressants, antichloesterolic, anticancer, antifungal etc. As a part of the ongoing study, we have synthesized some new benzothiazines, their sulfones and ribofuranosides. Substituted benzothiazines were prepared by condensation of 2-aminobenzenethiols with β -diketone/ β ketoester in presence of dimethylsulfoxide through oxidative cyclization. Intermediate bis-(2-aminophenyl)disulfides undergoes cyclization through scission of S-S bond due to high reactivity of alpha position of enaminoketone system towards nucleophilic attack. The structures of these compounds were determined on the basis of spectral data and elemental analysis. These compounds were also screened for biological activity [1-11].

2. Experimental

All the melting points were determined in open capillary tubes but are uncorrected. ¹H NMR and ¹³C NMR were recorded on JEOL AL 300 spectrometer (300 MHz) in DMSO- d_6 / CDCl₃ using TMS (tetramethyl silane) as an internal standard (Chemical shifts are measured in δ , ppm). IR spectra were recorded in KBr on SHIMADZU 8400 S FTIR spectro-photometer. Mass spectra were recorded on JEOL SX 102/DA 600 using Xenon/Argon as FAB (Fast Atom Bombardment) gas. The purity of compounds were checked by thin layer chromatography using silica gel "G" as adsorbent, visualizing these by UV light or in an Iodine chamber. Elemental analysis of these compounds was also done.

ABSTRACT

Synthesis of heterocyclic compounds like benzothiazines has attracted attention in recent years due to their biological and industrial value. This article reflects up-to-date and comprehensive coverage of biochemical aspects of benzothiazines, their sulfones and ribofuranosides. The nitrogen and sulfur containing heterocycles were prepared by condensation followed by oxidative cyclization of 2-aminobenzenethiol with β -diketones/ β -ketoesters in dimethylsulfoxide. These compounds were then used as base to prepare ribofuranosides by treating them with sugar (β -D-ribofuranose-1-acetate-2,3,5-tribenzoate). On refluxing with hydrogen peroxide in glacial acetic acid, these substituted dimethyl 4*H*-1,4-benzothiazines yielded 4*H*-1,4-benzothiazine-1,1-dioxides. Antioxidant and antimicrobial activity of these compounds were carried out and structure evaluation was done by spectral and elemental analysis.

2.1. General method of synthesis of substituted 4H-1,4benzothiazine (3a-d)

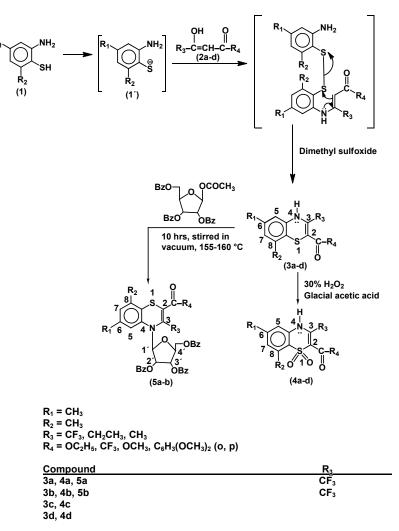
To a stirred suspension of 0.01 mole of β -diketone/ β -ketoester (**2a-d**) in 5 mL of dimethylsulfoxide was added 0.01 mole of 2-aminobenzenethiol (**1**) and resulting mixture was refluxed for 20 minutes (Table 1). The reaction mixture was concentrated, cooled down to room temperature. The solid separated out was filtered, washed with petroleum ether and crystallized from methanol (Scheme 1). Characterization data and spectral analysis of each compound (**3a-d**) is given as:

Ethyl-3-trifluoromethyl-6,8-dimethyl-4H-1,4-benzothiazine-2-carboxylate (**3a**): Yield: 46%. M.p.: 55 °C. IR (KBr, *v*, cm⁻¹): 3260 (N-H), 1690 (>C=O), 1340, 1160 (CF₃ str.), 1255, 1015 (C-O-C str.), 2885 (-CH₃ str.). ¹H NMR (DMSO-*d*₆, 300 MHz, δ , ppm): 8.90 (s, 1H, N-H), 8.08-7.20 (m, 2H, aromatic-H), 4.19 (q, 2H, *J* = 6.9 Hz, CH₂ of C₂H₅ at C₂), 1.30 (t, 3H, *J* = 6.2 Hz, CH₃ of C₂H₅ at C₂), 2.35 (s, 3H, CH₃ at C₆), 2.36 (s, 3H, CH₃ at C₈). ¹³C NMR (CDCl₃, 300 MHz, δ , ppm): 107.2 (C-2), 138.6 (C-3), 113.1 (C-5), 135.2 (C-6), 120.2 (C-7), 139.1 (C-8), 21.2 (CH₃ at C₆), 14.1 (CH₃ at C₈), 165 (C of CO at C₂), 114 (-CF₃ at C₃), 59.2 (CH₂ of COOC₂H₅ at C₂), 13.8 (CH₃ of COOC₂H₅ at C₂), 13.8 (CH₃ of COOC₂H₅ at C₂), 13.8 (CH₃ of COOC₂H₅ at C₂), 13.7 (M⁺), 244 (52), 202 (38), 275 (76), 73 (100). Anal. calcd. for C₁₄₁₄₄N0₂F₃S.

2-Trifluoroacetyl-3-trifluoromethyl-6,8-dimethyl-4H-1,4benzothiazine (**3b**): Yield: 68%. M.p.: 65 °C. IR (KBr, v, cm⁻¹): 3385 (N-H), 1650 (>C=O), 1350, 1180 (CF₃ str.), 2895 (-CH₃ str.). ¹H NMR (DMSO- d_6 , 300 MHz, δ , ppm): 8.29 (s, 1H, NH), 8.26-6.06 (m, 2H, aromatic-H), 2.35 (s, 3H, CH₃ at C₆), 2.36 (s, 3H, -CH₃ at C₈). ¹³C NMR (CDCl₃, 300 MHz, δ , ppm): 116.2 (C-2), 136.3 (C-3), 112.2 (C-5), 136.4 (C-6), 119.2 (C-7), 138.2 (C-8), 15.8 (CH₃ at C₆), 20.1 (CH₃ at C₈), 196.5 (C of CO at C₂), 129.8 (CF₃ at C₂), 114.1 (CF₃ at C₃).

 Table 1. The exact reaction times and yields of individual reactions (3a-d, 4a-d, 5a-b).

Compound No	R_3	R4	Reaction times (min.)	Yield (%)
3a	CF ₃	OC ₂ H ₅	22	46
3b	CF ₃	CF ₃	20	68
3c	CH_2CH_3	OCH ₃	23	76
3d	CH ₃	$C_6H_3(OCH_3)_2(o, p)$	21	80
4a	CF ₃	OC ₂ H ₅	255	69
4b	CF ₃	CF ₃	275	72
4c	CH ₂ CH ₃	OCH ₃	295	56
4d	CH_3	$C_6H_3(OCH_3)_2(o, p)$	260	82
5a	CF ₃	OC ₂ H ₅	920	84
5b	CF ₃	CF ₃	980	68



Scheme 1

MS (*m*/*z*, %): 341 (M⁺), 244 (58), 202 (29), 327 (78), 97 (100). Anal. calcd. for C₁₃H₉NOF₆S: C, 45.74; H, 2.63; N, 4.10. Found: C, 45.98; H, 2.65; N, 4.06%.

Methyl-3-ethyl-6,8-dimethyl-4H-1,4-benzothiazine-2carboxylate (**3c**): Yield: 76%. M.p.: 115 °C. IR (KBr, *ν*, cm⁻¹): 3280 (N-H), 1700 (>C=O), 1240, 1055 (C-O-C str.), 2875 (-CH₃ str.). ¹H NMR (DMSO-*d*₆, 300 MHz, δ, ppm): 8.40 (s, 1H, NH), 7.29-6.38 (m, 2H, aromatic-H), 2.35 (s, 3H, -CH₃ at C₆), 2.36 (s, 3H, -CH₃ at C₈), 2.00 (q, 2H, *J* = 7.01 Hz, CH₂ of CH₂CH₃ at C₃), 1.06 (t, 3H, *J* = 6 Hz, CH₃ of CH₂CH₃ at C₃), 3.76 (s, 3H, -CH₃ of CH₂3 at C₂). ¹³C NMR (CDCl₃, 300 MHz, δ, ppm): 108.2 (C-2), 141.7 (C-3), 118.2 (C-5), 139.1 (C-6), 114.2 (C-7), 142.2 (C-8), 19.6 (CH₃ at C₆), 14.8 (CH₃ at C₈), 24.1 (CH₂ of CH₂CH₃ at C₃), 8.5 (CH₃ of CH₂CH₃ at C₃), 16.0 (C of CO at C₂), 50.1 (C of OCH₃ at C₂). MS (*m*/*z*, %): 263 (M⁺), 204 (56), 162 (31), 221 (71), 59 (100). Anal. calcd. for C₁₄H₁₇NO₂S: C, 63.87; H, 6.46; N, 5.32. Found: C, 64.11; H, 6.40; N, 5.39%.

2-(2',4'-Dimethoxybenzoyl)-3,6,8-trimethyl-4H-1,4-benzothiazine (**3d**): Yield: 80%. M.p.: 53 °C. IR (KBr, v, cm⁻¹): 3298 (N-H), 1690 (>C=0), 1260, 1060 (C-O-C str.), 2886 (CH₃ str.). ¹H NMR (DMSO- d_6 , 300 MHz, δ , ppm): 9.09 (s, 1H, NH), 7.16-6.08 (m, 2H, aromatic-H), 1.71 (s, 3H, -CH₃ at C₃), 3.73 (s, 3H, -OCH₃ at ortho position of -COC₆H₃ (OCH₃)₂ (o,p) at C₂), 3.78 (s, 3H, -OCH₃ at para position of -COC₆H₃ (OCH₃)₂ (o,p) at C₂), 2.14 (s, 3H, CH₃ at C₆), 2.20 (s, 3H, CH₃ at C₈), 7.20-5.95 (m, 3H, aromatic-H of benzoyl group). ¹³C NMR (CDCl₃, 300 MHz, δ , ppm): 112.9 (C-2), 138.6 (C-3), 111.9 (C-5), 136.2 (C-6), 118.2 (C-7), 146.2 (C-8), 16.5 (CH₃ at C₃), 187 (C of CO at C₂), 56.8 (C of OCH₃ at ortho position $-COC_6H_3$ (OCH₃)₂ (o, p) at C₂). MS (*m/z*, %): 355 (M⁺), 190 (61), 137 (48), 148 (28), 313 (70), 165 (100). Anal. calcd. for C₂₀H₂₁NO₃S: C, 67.60; H, 5.91; N, 3.94. Found: C, 67.88; H, 5.89; N, 3.89%.

2.2. General method of synthesis of 4H-1,4-benzothiazine, 1,1-dioxides (sulfones) (4a-d)

30% Hydrogen peroxide (5 mL) was added to a solution of 0.01 mole of 4*H*-1,4-benzothiazine in 20 mL glacial acetic acid and refluxed for 15 minutes at 50-55 °C. Heating was stopped and another lot of 5 mL of 30%. Hydrogen peroxide was added. The reaction mixture was again refluxed for 4-5 hrs. The excess of solvent was removed by distillation under reduced pressure and the solution was poured into a beaker containing crushed ice. The yellow residue separated out was filtered and then crystallized from ethanol (Scheme 1). Characterization and spectral data of these compounds (**4a-d**) is given as:

Ethyl-3-trifluoromethyl-6,8-dimethyl-4H-1,4-benzothiazine-2-carboxylate-1,1-dioxide (**4a**): Yield: 69%. M.p.: 70 °C. IR (KBr, *v*, cm⁻¹): 3270 (N-H), 1700 (>C=0), 1180, 1140 (SO₂ sym. str.), 1080 (C-S str.). ¹H NMR (DMSO-*d₆*, 300 MHz, δ , ppm): 8.95 (s, 1H, NH), 8.06-7.30 (m, 2H, aromatic-H), 4.20 (q, *J* = 6.95 Hz, 2H, CH₂ of C₂H₅ a C₂), 1.29 (t, 3H, *J* = 6.3 Hz, CH₃ of C₂H₅ at C₂), 2.35 (s, 3H, -CH₃ at C₆), 2.36 (s, 3H, -CH₃ at C₈). ¹³C NMR (CDCl₃, 300 MHz, δ , ppm): 96.2 (C-2), 150.1 (C-3), 114.9 (C-5), 143.4 (C-6), 120.9 (C-7), 136.4 (C-8), 114.5 (-CF₃ at C₃), 168 (C of CO of COOC₂H₅ at C₂), 59.6 (CH₂ of COOC₂H₅ at C₂), 13.7 (CH₃ of COOC₂H₅ at C₂). MS (*m*/*z*, %): 349 (M⁺), 276 (51), 234 (29), 307 (75), 73 (100). Anal. calcd. for C₁₄H₁₄NO₄F₃S: C, 48.13; H, 4.01; N, 4.01. Found: C, 48.35; H, 4.05; N, 4.03%.

2-Trifluoroaceyl-3-trifluoromethyl-6,8-dimethyl-4H-1,4benzothiazine--1,1-dioxide (**4b**): Yield: 72%. M.p.: 220 °C. IR (KBr, v, cm⁻¹): 3390 (N-H), 1680 (>C=O), 1172, 1150 (SO₂ sym. str.), 1085 (C-S str.). ¹H NMR (DMSO-d₆, 300 MHz, δ , ppm): 8.30 (s, 1H, NH), 8.16-7.08 (m, 2H, aromatic-H), 2.35 (s, 3H, -CH₃ at C₆), 2.36 (s, 3H, -CH₃ at C₈). ¹³C NMR (CDCl₃, 300 MHz, δ , ppm): 101 (C-2), 154 (C-3), 118 (C-5), 144 (C-6), 121 (C-7), 146 (C-8), 21.2 (CH₃ at C₆), 13.7 (-CH₃ at C₃), 114 (-CF₃ at C₃), 196.8 (C of CO at C₂), 128.8 (CF₃ at C₂). MS (*m*/*z*, %): 373 (M⁺), 276 (48), 234 (28), 331 (72), 97 (100). Anal. calcd. for C₁₃H₉NO₃F₆S: C, 41.82; H, 2.41; N, 3.75. Found: C, 42.09; H, 2.39; N, 3.78%.

Methyl-3-Ethyl-6,8-dimethyl-4H-1,4-benzothiazine-2-carboxylate-1,1-dioxide (**4c**): Yield: 56%. M.p.: 240 °C. IR (KBr, v, cm⁻¹): 3290 (N-H), 1710 (>C=O), 1165, 1160 (SO₂ sym. str.), 1095 (C-S str.). ¹H NMR (DMSO- d_6 , 300 MHz, δ , ppm): 8.80 (s, 1H, NH), 7.30-6.40 (m, 2H, aromatic-H), 2.16 (q, 2H, *J* = 7.2 Hz, CH₂ of CH₂CH₃ at C₃), 1.09 (t, 3H, *J* = 6.4 Hz, CH₃ of CH₂CH₃ at C₃), 3.79 (s, 3H, -CH₃ of OCH₃ at C₂), 1.86 (s, 3H, CH₃ at C₆). 2.31 (s, 3H, CH₃ at C₈). ¹3C NMR (CDCl₃, 300 MHz, δ , ppm): 99.2 (C-2), 144 (C-3), 113.8 (C-5), 145.1 (C-6), 120.9 (C-7), 143 (C-8), 25.2 (CH₂ Of CH₂CH₃ at C₃), 8.9 (CH₃ at CH₂CH₃ at C₃). MS (*m/z*, %): 295 (M⁺), 236 (38), 194 (30), 253 (76), 59 (100). Anal. calcd. for C₁₄H₁₇No₄S: C, 56.94; H, 5.76; N, 4.74. Found: C, 57.22; H, 5.70; N, 4.78%.

2-(2',4'-Dimethoxybenzoyl)-3,6-8-trimethyl-4H-1,4-benzothiazine-1,1-dioxide (4d): Yield: 82%. M.p.: 81 °C. IR (KBr, v, cm⁻¹): 3320 (N-H), 1700 (>C=O), 1177, 1150 (SO₂ sym. str.), 1077 (C-S str.). ¹H NMR (DMSO-*d*₆, 300 MHz, δ , ppm): 9.12 (s, 1H, NH), 7.23-6.09 (m, 2H, aromatic-H), 1.92 (s, 3H, -CH₃ at C₃), 3.92 (s, 3H, -OCH₃ at ortho position of -COC₆H₃(OCH₃)₂ (o, p) at C₂), 3.79 (s, 3H, -OCH₃ at para position of COC₆H₃(OCH₃)₂ (o, p) at C₂). ¹3C NMR (CDCl₃, 300 MHz, δ , ppm): 97 (C-2), 153 (C-3), 113.8 (C-5), 138 (C-6), 123 (C-7), 145 (C-8), 16.2 (CH₃ at C₃), 189 (C of CO c6₆H₃(OCH₃)₂ (o, p) at C₂). 56.9 (C of OCC₆H₃(OCH₃)₂ (o, p) at C₂). MS (*m*/*z*, %): 387 (M⁺), 222 (50), 180 (21), 345 (70), 165 (100). Anal. calcd. for C₂₀H₂₁NO₅S: C, 62.01; H, 5.42; N, 3.61. Found: C, 62.26; H, 5.38; N, 3.56%.

2.3. General method of synthesis of substituted N-(2',3',5'-tri-O-benzoyl- β -D-ribofuranosyl)benzothiazine (5a-b)

To a concentrated solution of synthesized benzothiazines (**3a-d**), (0.002) mole in toluene, β -D-ribofuranose-1-acetate-2,3,5-tribenzoate (0.002) mole was added and stirred, in vacuuo, on an oil bath, at 155-160 °C, for 15 minutes. The vacuum was broken and the reaction was protected from moisture by using a guard tube. Stirring was further continued for 10-15 hours with application of vacuum for 15 minutes after every hour. The melt was dissolved in methanol, boiled for 10 minutes and cooled to room temperature. The precipitate was filtered and the filtrate was evaporated to dryness. The viscous residue, thus obtained was dissolved in ether, filtered, concentrated and kept in refrigerator overnight to get the crystalline compound (**5a-b**) (Scheme 1). Characterization and spectral data of these compounds (**5a-b**) is given as:

N-(2',3',5'-tri-O-benzoyl-β-D-ribofuranosyl)-ethyl-3-trifluoro methyl-6,8-dimethyl-4H-1,4-benzothiazine-2-carboxylate (5a): Yield: 84%. M.p.: 80 °C. IR (KBr, v, cm⁻¹): 1700 (C=O), 1350, 1170 (-CF₃ str.), 1170 (C-O-C str.). ¹H NMR (DMSO- d_6 , 300 MHz, δ , ppm): 8.52-7.10 (m, 2H, aromatic-H), 4.21 (q, 2H, *J* = 6.8 Hz, CH₂ of C₂H₅ at C₂), 1.40 (t, 3H, *J* = 6.86 Hz, 'CH₃ of C₂H₅ at C₂). ¹³C NMR (CDCl₃, 300 MHz, δ , ppm): 108.2 (C-2), 139 (C-3), 113.2 (C-5), 136.8 (C-6), 121.2 (C-7), 140.2 (C-8), 66.8 (C-1'), 78.5 (C-2'), 72.5 (C-3'), 71.6 (C-4'). MS (*m*/*z*, %): 761 (M⁺), 688 (39), 646 (31), 73 (100). Anal. calcd. for C₄₀H₃₄NO₉F₃S: C, 63.07; H, 4.46; N, 1.83. Found: C, 63.28; H, 4.40; N, 1.80%. Optical rotation [α]_b 23= -17.28 °C.

N-(2['],3['],5[']-Tri-O-benzoyl-β-D-ribofuranosyl)-2-trifluoro acetyl-3-trifluoromethyl-6,8-dimethyl-4H-1,4-benzothiazine (**5b**): Yield: 68%. M.p.: 82 °C. IR (KBr, v, cm⁻¹): 1680 (C=O), 1355, 1185 (-CF₃ str.), 1165 (C-O-C str.). ¹H NMR (DMSO- d_6 , 300 MHz, δ , ppm): 8.28-7.01 (m, 2H, aromatic-H), 2.38 (s, 2H, CH₃ at C₆), 2.37 (s, 3H, -CH₃ at C₈). ¹³C NMR (CDCl₃, 300 MHz, δ , ppm): 117.2 (C-2), 173.3 (C-3), 113.6 (C-5), 136.4 (C-6), 120 (C-7), 136 (C-8), 69.2 (C-1), 75.2 (C-2), 76.1 (C-3'), 72.1 (C-4'). MS (m/z, %): 785 (M⁺), 688 (37), 646 (27), 97 (100). Anal. calcd. for C₃₉H₂₉NO₈F₆S: C, 59.61; H, 3.69; N, 1.78. Found: C, 59.83; H, 3.63; N, 1.76%. Optical rotation [α]₂ 23 = -20.18 °C.

2.4. Biological activity

2.4.1. Antioxidant activity

2.4.1.1. DPPH radical scavenging assay

Radical scavenging activity of the synthesized compounds against stable 1,1-diphenyl-2-picrylhydrazyl (DPPH) radical was determined spectrophotometrically as described by Cuendet *et al.* [12]. A stock solution of 1 mg/mL of the compound was prepared in methanol. 50 μ L of compounds were added to 5 mL of a 0.004% methanol solution of DPPH. After 30 minutes incubation in dark at room temperature, absorbance was read against a blank at 517 nm (Table 2). Since IC50 Value is inversely related to the antioxidant activity; ascorbic acid, which is a very good antioxidant, shows a lower IC50 value of 17.8 μ g/mL. Among all the synthesized compounds; compound **3c** shows a value of 23.56 μ g/mL which shows its good antioxidant nature.

The assay was carried out in triplicate and the percentage of inhibition was calculated by using the following formula.

% Inhibition =
$$\frac{(AB - AA)}{AB} \times 100$$
 (1)

where AB = Absorption of blank, AA = Absorption of test, Ascorbic acid (shows antioxidant activity) as a positive control and methanol (no antioxidant activity) as a negative control has been used in this assay.

Table 2. Antioxidant activity of the synthesized compounds (DPPH) assay (3a-d, 4a-d, 5a-b).

Compound No	R_3	R ₄	DPPH % inhibition of 1 mg/mL of compound	IC ₅₀ (μg/mL)
3a	CF ₃	OC ₂ H ₅	42.74 ± 0.03	823.78
3b	CF ₃	CF ₃	65.13 ± 0.02	500.8
3c	CH_2CH_3	OCH ₃	90.76 ± 0.07	23.56
3d	CH ₃	C ₆ H ₃ (OCH ₃) ₂ (o, p)	70.79 ± 0.08	79.12
4a	CF ₃	OC ₂ H ₅	23.48 ± 0.09	-
4b	CF ₃	CF ₃	67.49 ± 1.10	432.90
4c	CH ₂ CH ₃	OCH ₃	11.88 ± 0.05	-
4d	CH_3	$C_6H_3(OCH_3)_2(o, p)$	34.10 ± 1.20	-
5a	CF ₃	OC ₂ H ₅	73.27 ± 0.06	62.70
5b	CF ₃	CF ₃	75.37 ± 0.08	58.90
Ascorbic acid			92.96 ± 00.9	17.82

Table 3. Antioxidant activity f the synthesized compounds* (ABTS++ assay) (3a-d, 4a-d, 5a-b).

Compound No	R ₃	R ₄		ABTS ++ activity at different intervals (min.)				
			0	1	2	4	6	
3a	CF ₃	OC ₂ H ₅	0.697	0.696	0.691	0.687	0.482	
3b	CF ₃	CF ₃	0.698	0.303	0.264	0.097	0.025	
3c	CH ₂ CH ₃	OCH ₃	0.688	0.109	0.09	0.002	0.002	
3d	CH ₃	$C_6H_3(OCH_3)_2(o, p)$	0.695	0.087	0.039	0.025	0.025	
4a	CF ₃	OC ₂ H ₅	0.699	0.525	0.406	0.365	0.339	
4b	CF ₃	CF ₃	0.686	0.392	0.367	0.294	0.262	
4c	CH ₂ CH ₃	OCH ₃	0.693	0.682	0.609	0.382	0.329	
4d	CH ₃	$C_6H_3(OCH_3)_2(o, p)$	0.700	0.276	0.162	0.027	0.021	
5a	CF ₃	OC ₂ H ₅	0.686	0.194	0.095	0.032	0.016	
5b	CF ₃	CF ₃	0.714	0.228	0.195	0.069	0.023	
Ascorbic acid			0.694	0.040	0.003	0.003	0.003	

*Ascorbic acid is used as a reference compound.

Table 4. Antimicrobial activity of the synthesized compounds (3a-d, 4a-d, 5a-b).

				Antifungal activity b		
Compound No	R ₃	R4	Coagulase negative staphylococci	Coagulase positive staphylococci	Enterobacter	Candida albicans
3a	CF ₃	OC ₂ H ₅	23	10	10	18
3b	CF ₃	CF ₃	11	11	13	22
3c	CH ₂ CH ₃	OCH ₃	15	13	-	18
3d	CH ₃	C ₆ H ₃ (OCH ₃) ₂ (o, p)	16	12	12	15
4a	CF ₃	OC ₂ H ₅	12	10	-	-
4b	CF ₃	CF ₃	11	10	11	11
4c	CH ₂ CH ₃	OCH ₃	-	-	-	11
4d	CH ₃	C ₆ H ₃ (OCH ₃) ₂ (o, p)	10	11	-	18
5a	CF ₃	OC ₂ H ₅	10	-	11	14
5b	CF ₃	CF ₃	20	10	-	12
Gatifloxacin			-	-	17	-
Vancomycin			15	15	-	-
Flucanazole			-	-	-	25

^a Zone of inhibition in mm; <7 mm inactive; 7-9 mm weakly active; 10-12 mm, moderately active; >12 mm, active.

^b Zone of inhibition in mm; <7 mm inactive; 7-11 mm weakly active; 12-17 mm, moderately active; >17 mm, active.

2.4.1.2. ABTS radical cation decolorization assay

The 2,2-azinobis(3-ethybenzothiazoline-6-sulphonic acid) radical cation (ABTS⁺⁺) decolorization test was carried out using an improved assay of Re *et al.* [13]. In brief, ABTS⁺⁺ was generated by oxidation of ABTS with potassium persulphate. For this purpose, ABTS was dissolved in ionized water at concentration of 7 mM, and potassium persulphate was added to a concentration of 2.45 mM. The reaction mixture was left at room temperature overnight (12-15 hours), in the dark before use; the ABTS solution then was diluted with ethanol to an absorbance of 0.700 ± 0.020 at 734 nm. After addition of 1 ml of the diluted ABTS solution to 10 μ L of compound and mixing, absorbance readings were taken at 30 °C at intervals of exactly 1-6 min. The experiment was carried out in triplicate (Table 3 and Figure 1). We have used ascorbic acid as a positive control and ethanol as a negative control in this assay.

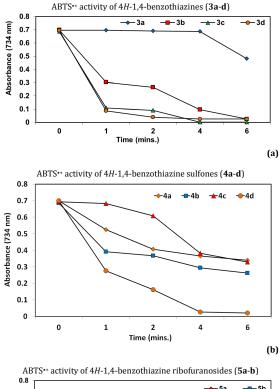
2.4.2. Antimicrobial activity

The antimicrobial assay of the synthesized compounds was carried out by using paper disc method of Gould *et al.* [13] against some bacteria and fungi at 100 μ g per disc concentration using Vancomycin, Gatifloxacin as reference compounds against bacteria (*Coagulase negative Staphylococci, Coagulase positive Staphylococci, Enterobacter*) and Flucanazole against fungus (*Candida albicans*). Paper disc method includes

preparation of plates by pouring molten media into sterile petriplates which was then allowed to solidify for 5 minutes and 0.1% inoculum suspension was swabbed uniformly and allowed to dry for 5 min. The compound discs prepared were then placed over the plates and incubated for $37 \circ C$ for 24 hrs. At the end inhibition zones were measured with ruler in millimetre. These microorganisms were obtained from Microbiology Department, Swai Man Singh Medical College, Jaipur (Table 4).

3. Results and discussion

2-Aminobenzenethiol (1) and β -diketones / β -ketoesters (2a-d) were refluxed in dimethyl sulfoxide which involves condensation and oxidative cyclization. A *bis*-(2-aminophenyl) disulfide (1) was obtained by oxidation of 2-aminobenzene thiol which cyclizes to form 4*H*-1,4-benzothiazines (3a-d) by cleavage of sulfur-sulfur bond due to high reactivity of α -position of enaminoketone system towards nucleophillic attack. Compound (3a-d) on treatment with 30% hydrogen peroxide in glacial acetic acid were converted into their corresponding sulfones (4a-d). Treatment of (3a-b) in toluene with β -D-ribofuranose-1-acetate-2,3,5-tribenzoate in vacuum gave the corresponding ribofuranosides (5a-b) (Scheme 1). The structures of synthesized compounds are well supported by spectral data and elemental analysis.



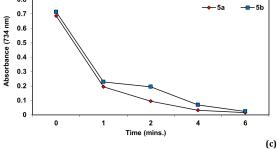


Figure 1. The effect of time on the suppression of absorbance of ABTS by synthesized compounds. After addition of 1 mL of diluted ABTS solution (A 734 nm = 0.700 ± 0.020) to 10 µL of the compound the absorbance reading was taken at 30 °C exactly 1 min., after initial mixing and up to 6 min. All determinations were carried out in triplicates.

3.1. Spectral analysis

3.1.1. IR spectra

Compounds (**3a-d**) showed peaks in region 3385-3260 cm⁻¹ due to N-H stretching vibrations and 1700-1650 cm⁻¹ due to >C=O stretching vibrations which gets shifted to higher frequencies to 3390-3270 cm⁻¹ and 1710-1680 cm⁻¹, respectively, in compounds (**4a-d**). Compounds (**4a-d**) also exhibited two intense peaks in region 1360-1340 cm⁻¹ and 1180-1140 cm⁻¹ due to asymmetric and symmetric stretching vibrations of sulfonyl group. Compounds (**4a-d**) also showed C-S stretching vibrations in region 1095-1077 cm⁻¹.

Absence of stretching vibrations due to >N-H group in compounds (**5a-b**) showed site of ribosylation, further in compounds (**5a-b**) bands due to C-O-C linkage of sugar appeared in the region 1170-1165 cm⁻¹.

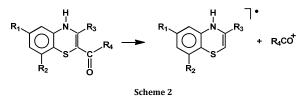
3.1.2. ¹H NMR spectra

All compounds showed multiplet in region δ 8.52-6.06 ppm due to aromatic protons and compounds (**3a-d**) and (**4a-d**) also

showed a singlet due to N-H proton in the region δ 9.12-8.29 ppm. Peak due to N-H proton was found to be absent in compounds (**5a-b**) due to ribosylation. In ribofuranosides, C'₄-H proton showed multiplet in region δ 4.43-4.80 ppm, C₂'-H and C₃'-H protons appeared in region δ 4.51-5.90 ppm as multiplet and C₁'-H proton appeared as doublet at δ 6.30-6.40 ppm.

3.1.3. Mass spectra

The molecular ion peaks of 4H-1,4-benzothiazines were in accordance with their molecular weights. In all the cases side chain at C₂ appears as a base peak which is obtained by its fission (Scheme 2).



3.2. Biological activity (Antioxidant and Antimicrobial)

All the synthesized compounds (**3a-d**), (**4a-d**), and (**5a-b**) were screened for their antioxidant activity by DPPH radical scavenging assay and (ABTS^{•+}) radical cation decolorization assay. The synthesized compounds were also screened for antimicrobial activity (antibacterial and antifungal) by paper disc method. The present study demonstrated that these compounds showed mixed activity in DPPH and ABTS^{•+} assay.

(1) Compounds (**3b**, **3c**, **3d**, **4b**, **5a**, **5b**) showed strong radical scavenging activity in DPPH assay that have DPPH % inhibition \geq 50.

(2) Compounds (**3a**, **4d**) showed moderate activity in DPPH assay that have DPPH % inhibition \ge 30.

(3) Compounds (4a, 4c) showed mild activity (<30) in DPPH assay.

(4) Compounds (**3b**, **3c**, **3d**, **4b**, **4d**, **5a**, **5b**) were active in ABTS⁺⁺ assay.

All these compounds were found to be moderately active against various bacteria such as (*Coagulase negative Staphylococci, Coagulase positive Staphylococci, Enterobacter*) and fungi (*Candida albicans*). Compounds (**3a, 3c, 3d, 5b**) showed good activity against *Coagulase negative Staphylococci* and compounds (**3a, 3b, 3c** and **4d**) shows good activity against *Candida albicans*.

4. Conclusion

The structures of synthesized compounds are well supported by spectral data and elemental analysis. The synthesized compounds were also screened for antioxidant activity (DPPH assay and ABTS⁺⁺ assay) and antimicrobial activity (antibacterial and antifungal). The present study demonstrated that these compounds showed mixed activity in DPPH and ABTS⁺⁺ assay. Compounds **3c**, **3d** showed excellent antioxidant activity in DPPH assay. Compounds **5a**, **5b** showed much better activity in DPPH assay than compounds **3a**, **3b** which are the precursors of **5a** and **5b**. It shows that the ribofuranosides **5a** and **5b** showed much better antioxidant activity than their phenothiazine bases (**3a**, **3b**). This is due to the replacement of H by the sugar moiety (ribosylation). Compounds **3b**, **3c**, **3d**, **4b**, **5a**, **5b** showed good activity in both DPPH and ABTS assays.

The present paper is focused on the synthesis of novel heterocyclic compounds as possible antibacterial and antifungal agents. Compounds **3c** and **3d** showed antibacterial activity against *Coagulase* negative *Staphylococci* which is comparable to vancomycin. Compound **3a** is much better than

vancomycin in antibacterial activity against *Coagulase* negative *Staphylococci*. All these compounds were found to be moderately active against various bacteria such as (*Coagulase negative Staphylococci, Coagulase positive Staphylococci, Enterobacter*) and fungi (*Candida albicans*). Compounds (**3a**, **3c**, **3d**, **5b**) showed good activity against *Coagulase negative Staphylococci* and compounds (**3a**, **3b**, **3c** and **4d**) shows good activity against *Candida albicans*.

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References

- [1]. Gautam, V.; Sharma, M.; Panwar, M.; Gautam, N.; Kumar, A.; Sharma, I. K.; Gautam, D. C. Phosphorus Sulfur **2009**, 184(11), 3090-3109.
- [2]. Gupta, S.; Ajmera, N.; Meena, P.; Gautam, N.; Kumar, A.; Gautam, D. C. Jordan J. Chem. 2009, 4(3), 209-221.
- [3]. Gupta, V.; Gautam, R. K.; Jain, S. K.; Gupta, R. R. Phosphorus Sulfur 1990, 47(1), 225-228.
- [4]. Clercq, E. D. Nucleos. Nucleot. Nucl. 1985, 4(1), 3-11.
 [5]. Gupta, A.; Saraswat, V.; Mukherji, S. K.; Gupta, R. R. Phosphorus Sulfur
- 1993, 85(1), 101-106.
 [6]. Kachee, T. L.; Gupta, V.; Gautam, D. C.; Gupta, R. R. Phosphorus Sulfur 2005, 180(10), 2225-2234.
- [7]. Gautam, N.; Hans, D.; Gautam, D. C. Oriental J. Chem. 2005, 21(2), 299-302.
- [8]. Kumar, N.; Singh, G.; Yadav, A. K. Heteroatom Chem. 2001, 12(1), 52-56.
- [9]. Gupta, R. R. Phenothiazines and 1,4-benzothiazines Chemical and Biomedical aspects, Elsevier, Amsterdam, 1988 pp. 160-210.
- [10]. Sharma, P. R.; Gupta, V.; Gautam, D. C.; Gupta, R. R. Phosphorus Sulfur 2003, 178(7), 1483-1488.
- [11]. Gautam, N.; Sharma, M.; Gautam, V. Asian J. Chem. 2010, 22(7), 5380-5388.
- [12]. Cuendet, M.; Hostettmann, K.; Potterat, O.; Dyatmiko, W. Helv. Chim. Acta 1997, 80(4), 1144-1152.
- [13]. Re, R.; Pellegrini, N.; Proteggente, A.; Pannala, A.; Yang, M.; Rice-Evans, C. Free Radic. Biol. Med. **1999**, 26, 1231-1237.
- [14]. Gould, J. C.; Browie, J. H. Edinb. Med. J. 1952, 59(4), 178-199.